

REVIEW ARTICLE | OPEN ACCESS

SAFE pathway and conditioning: from discovery to its effectiveness in the presence of cardiovascular risk factors

Aqeela Imamdin¹, Nkanyiso Hadebe^{1,2} and Sandrine Lecour¹

Although ischemic conditioning has been described as the most powerful strategy to protect against ischemia-reperfusion injury, its translation to the clinical setting has proved to be challenging, possibly due, at least in part, to the presence of different cardiovascular risk factors in patients, which are known to affect its efficacy. Ischemic conditioning confers cardioprotection via the activation of multiple prosurvival pathways, including the activation of the survivor activating factor enhancement (SAFE) path, which involves mediators of the innate immune system (i.e. tumor necrosis factor alpha) and key transcription factors such as the signal transducer and activator of transcription 3 (STAT3). Here, we describe the origin of the SAFE path as a key pathway in ischemic conditioning and we explore how the activation of this path may be affected in the presence of different cardiovascular risk factors.

Keywords: Prosurvival signalling pathways, SAFE, Tumor Necrosis Factor, signal transducer and activator of transcription 3, cardiovascular risk factors

The discovery of intrinsic cardioprotective signaling pathways in conditioning

Since its discovery in 1986, the phenomenon of ischemic conditioning has been the focus of intense research with more than 20,000 peer-reviewed publications (listed in PubMed) trying to understand this phenomenon and to translate it to the clinical arena (Cour and Lecour, 2019; Hausenloy et al., 2019). Unfortunately, as simple as ischemic conditioning may be applied (few episodes of ischemia/reperfusion prior to or after a sustained ischemic insult), these 34 years of research conducted worldwide have not been able to fully elucidate the mechanisms involved in this protective phenomenon, even though some important discoveries have been made over that time. In the 1990s, adenosine and protein kinase C were suggested as key components of the signaling cascade, their activation leading to the downstream activation of an unknown effector for cardioprotection (see Goto et al., 1996). The putative mitochondrial potassium adenosine triphosphate dependent channel was later proposed as the end effector of the cardioprotective signaling cascade, although the existence of this channel remained disputed for many years (Cohen et al., 2000; Paggio et al., 2019). The beginning of the 21st century was marked by research suggesting the involvement of the mitochondrial permeability transition pore (MPTP) whose identification has also revealed to be challenging (Downey

et al., 2007). The discovery of the reperfusion injury salvage kinase (RISK) pathway also represented a major advance in the understanding of the cardioprotective signaling pathways, with the activation of both protein kinase B (Akt) and extracellular signal-regulated kinase (ERK) at the onset of reperfusion having proved to play a key role to limit infarct size following an ischemic conditioning stimulus (Schulman et al., 2002). In 2009, we described the activation of the survivor activating factor enhancement (SAFE) pathway as an alternative path to the RISK pathway for cardioprotection (Lecour, 2009a). The SAFE path is triggered by the activation of the immune system with tumor necrosis factor-alpha (TNF) initiating the activation of a myriad of prosurvival components including signal transducer and activator of transcription 3 (STAT3) (Lecour, 2009b).

The SAFE pathway and its origins

The research that led to the discovery of the SAFE pathway goes back to 2000 when data published by Douglas Mann's group triggered our curiosity. Although TNF was known as a cytokine contributing to cardiac dysfunction in the context of heart failure, his group observed that mice with TNF receptor 1 (TNFR1) and TNF receptor 2 (TNFR2) deficiency presented with a larger left ventricular infarct size compared to their littermate controls when subjected to an ischemia-

¹Hatter Institute for Cardiovascular Research in Africa, Department of Medicine, Faculty of Health Sciences, University of Cape Town, South Africa.²Department of Anaesthesia, Faculty of Health Sciences, University of Cape Town, South Africa.

Correspondence should be addressed to Sandrine Lecour (Sandrine.lecour@uct.ac.za).

reperfusion injury (Kurrelmeyer et al., 2000). These data suggested that TNF may trigger some cardioprotective intrinsic signaling pathways and this led us to hypothesize that endogenous TNF may play an important role as a signaling component involved in the cardioprotective effect of ischemic conditioning. Using an isolated heart model, we discovered that TNF was able, in a dose dependent manner, to limit infarct size in hearts subjected to an ischemia-reperfusion injury when given as a preconditioning stimulus (Lecour et al., 2002). Similar findings were confirmed using an in vivo model of ischemia-reperfusion injury (Deuchar et al., 2007). Most importantly both ischemic pre- and post-conditioning failed to reduce the infarct size in mouse hearts from TNF knockout mice (Smith et al., 2002; Lacerda et al., 2009). When TNF was given as a preconditioning mimetic in mouse hearts, cardioprotection occurred in TNFR1 deficient mice but failed to protect in TNFR2 deficient mice, therefore suggesting that the cardioprotective effect offered with TNF is mediated via TNFR2 (Lacerda, 2009). Trying to delineate downstream targets of TNF, we found that both ischemic preconditioning and pharmacological conditioning with TNF increased the activation of STAT3 and failed to protect cardiomyocytes in STAT3 deficient mice (Lecour et al., 2005b; Suleman et al., 2008), therefore pinpointing STAT3 as a downstream target of TNF for cardioprotection. Although STAT3 was already known as a downstream target of interleukin 6, (another key component of the immune system) via the activation of glycoprotein 130, its activation by TNF and TNFR2 at the time was very intuitive (Kurdi and Booz, 2007). Today, the exact mechanisms by which TNF may activate STAT3 still remain to be elucidated. The dimerization (and therefore activation) of STAT3 is mediated via nicotinamide adenine dinucleotide dehydrogenase 1 alpha subcomplex 13 (NDUFA13), which is a subunit of the mitochondrial complex I (Hu et al., 2017). Most importantly, the activation of STAT3 downstream of TNFR2 activates optic atrophy 1 protein (OPA 1), which promotes mitochondrial fusion (Nan et al., 2017). Other downstream targets include the activation of protein kinase C, free radicals, the mitochondrial potassium ATP dependent channel, and the inhibition of the proapoptotic protein Bcl-2 antagonist of cell death (BAD)

(Lecour et al., 2002; Lecour et al., 2005a; Boengler et al., 2010; Boengler et al., 2013). Activated STAT3 translocates to the mitochondria and inhibits the opening of the MPTP, thus preventing mitochondrial leakage, and thereby improving the outcome of ischemia-reperfusion injury by maintaining cellular energy functions (Boengler et al., 2010). Very recently, we have also identified microRNAs as downstream targets of STAT3 for cardioprotection. Indeed, a microarray analysis performed in both control and STAT3 knockout mice suggested that miR-34b and miR-337 are downregulated downstream of STAT3 (Pedretti et al., 2019). Furthermore, in isolated cardiomyocytes, the expression of miR-34b and miR-337 was increased by hypoxia in isolated cardiomyocytes and reduced by cardioprotective strategies. In cardiomyocytes transfected with miRNA mimics, cardioprotective strategies failed to improve cell viability against hypoxia (Pedretti et al., 2019).

The activation of the SAFE path has been identified with different forms of conditioning including ischemic preconditioning (Lecour et al., 2005b), ischemic postconditioning (Lacerda et al., 2009), and remote ischemic conditioning (Tamareille et al., 2011). Crosstalk between the SAFE and the RISK paths has been suggested. In rodents subjected to an ischemia-reperfusion insult, a conditioning protocol failed to activate the key components of the RISK pathway Akt or ERK in the presence of a STAT3 inhibitor (Suleman et al., 2008; Tamareille et al., 2011; Somers et al., 2012). In a similar manner, inhibitors of Akt that blocked signling through the RISK pathway abolished the protection of an ischemic conditioning protocol, and this was associated with a failure to activate STAT3 (Suleman et al., 2008; Tamareille et al., 2011; Somers et al., 2012).

A particularity of the SAFE path compared to the RISK path is that it is activated both during the conditioning stimulus and at the onset of reperfusion following a conditioning protocol. Inhibitors of STAT3 activation such as AG490 abolished the cardioprotective effect of ischemic preconditioning when given either during the preconditioning stimulus (Suleman et al., 2008) or at the onset of reperfusion (Lecour et al., 2005b). Furthermore, STAT3 activation was observed at both time points (Lecour et al., 2005b; Suleman et al., 2008).



Figure 1. Cardiovascular risk factors such as age, gender, diabetes, and hypertension may affect the benefit of cardioprotective strategies by modulating the activation of the SAFE pathway. TNF: tumor necrosis factor alpha; STAT3: signal transducer and activator of transcription 3.

Table 1: Different conditioning stimuli confer cardioprotection via the activation of the survivor activating factor enhancement (SAFE) pathway. This path involves the activation of tumor necrosis factor alpha (TNF) and the signal transducer and activator of transcription 3 (STAT3) to promote cell survival.

Stimulus	Reference
Ischemic pre-/per-/post-conditioning	Lecour et al., 2005; Lacerda et al., 2009; Tamareilleet al., 2011
Remote ischemic conditioning	Tamareille et al., 2011
Insulin & leptin	Fuglesteg et al., 2008; Amantea et al., 2011
Erythropoietin	Watson et al., 2013
Oxytocin	Poshekan et al., 2018
Melatonin	Lamont et al., 2011
Exercise conditioning	Sun et al., 2018
HDL/ApoA /S1P	Kalakech et al., 2014; Frias et al., 2015
Hypothermia	Park et al., 2013
Anaesthetics	Soh et al., 2018
Glyceryl-trinitrate & cariporide	Kwan et al., 2015
Hydrogen sulphide	Calvert et al., 2009

HDL: High density lipoprotein; ApoA: Apolipoprotein A; S1P: Sphingosine-1 phosphate; GH: Growth hormone.

Do all conditioning strategies activate the SAFE pathway?

The cardioprotective effect of ischemic preconditioning, ischemic postconditioning, and remote conditioning have all been identified as being dependent on the activation of the SAFE pathway (Lecour 2005; Lacerda 2009; Tamareille 2011). Similarly, multiple pharmacological pre- or post-conditioning drugs activate the SAFE pathway for cardioprotection and these include: glyceryl trinitrate, cariporide, growth hormones release hormone, erythropoietin, oxytocin, hydrogen sulphide, high density lipoproteins, apolipoprotein A, sphingosine-1 phosphate, anaesthetics, melatonin, and resveratrol (Lecour et al., 2005b; Fuglesteg et al., 2008; Calvert et al., 2009; Lacerda et al., 2009; Amantea et al., 2011; Lamont et al., 2011; Tamareille et al., 2011; Frias et al., 2012; Park et al., 2013; Penna et al., 2013; Watson et al., 2013; Kalakech et al., 2014; Brulhart-Meynet et al., 2015; Kwan et al., 2015; Sun and Mao, 2018) (see figure 1). Conditioning with exercise or hypothermia also activates the SAFE path (Park et al., 2013; Sun and Mao, 2018).

Does the SAFE pathway offer multi-organ protection?

Although initially discovered to protect against cardiac ischemia-reperfusion injury, there is clear evidence that activation of the SAFE path to promote cell survival can also occur in other organs such as the brain (Wang et al., 2010; Sakata et al., 2012; Niemi et al., 2016), liver (Lee et al., 2016), lungs (Luo et al., 2018), hindlimbs (Han et al., 2016), and kidney (Dube et al., 2017). Surprisingly, remote liver ischemic preconditioning protects against cerebral ischemia-reperfusion injury via mechanisms, which may be independent of STAT3 activation (Yang et al., 2020).

Do traditional risk factors affect the effectiveness of the SAFE pathway?

Ischemic heart disease is often caused by or is associated with multiple risk factors including age, gender, hypertension, and diabetes. Most of these risk factors are known to alter many intrinsic cardiac signaling pathways in both physiological and pathophysiological conditions, and multiple studies suggest that they negatively interfere with the effectiveness of ischemic conditioning, most likely by modifying the sensitivity of the activation of prosurvival signaling pathways (see review by Ferdinandy et al., 2014). There is now strong evidence in the literature supporting that the SAFE pathway is also affected by these risk factors.

Age

Experimental data support evidence that aging affects the efficacy of conditioning with a lack of protection observed with ischemic postconditioning and remote ischemic conditioning in older rodents (Somers et al., 2011; Adam et al., 2013; Behmenburg et al., 2017; Heinen et al., 2018). TNF-mediated cardioprotective signalling pathways are impaired with aging (Cai et al., 2003). Most importantly, Boengler and colleagues reported that the lack of protection with ischemic postconditioning, observed in aged mice compared to younger animals, was associated with a lack of phosphorylation of STAT3 (Boengler et al., 2008). These data strongly suggest that reduced activation of the SAFE path with aging may contribute to the lack of protection with conditioning in aged mice.

Gender

Female and male hearts present a difference in susceptibility against ischemia-reperfusion injury with smaller infarcts observed in female compared to male when subjected to the same ischemia-reperfusion insult (Ostadal et al., 2009; Penna et al., 2009). Gender may also affect the effectiveness of conditioning. The benefit of ischemic postconditioning is larger in males compared to females, an effect which might be explained by the fact that the females were already better protected without the conditioning protocol (Penna et al., 2009). It is highly possible that some of these gender differences may be explained by a gender difference in sensitivity of the activation of the SAFE path. Indeed, gender differences in TNF signaling after ischemia/reperfusion have been reported, including a greater activation of TNFR2 in females compared to males (Wang et al., 2006; Wang et al., 2008). Similarly, male hearts subjected to an ischemia-reperfusion injury expressed a lower level of activated STAT3 compared to female hearts, an effect which was reversed in castrated males (Wang et al., 2009).

Diabetes

In both type 1 and type 2 diabetic animals, an increase in infarct size following an ischemia-reperfusion injury was observed, most likely as a consequence to an increase in hyperglycemia induced oxidative stress associated with the disease (see review (Lejay et al., 2016)). With no surprise, the cardioprotective effect of ischemic preconditioning and ischemic postconditioning was also compromised in diabetic animals (Przyklenk et al., 2011; Tyagi et al., 2019). The increased susceptibility of ischemia-reperfusion injury in the diabetic rats was associated with a decrease in STAT3 activation (Li et al., 2013). Similarly, STAT3 failed to be phosphorylated following a pharmacological or an ischemic conditioning stimulus in streptozotocin-induced diabetic rats (Li et al., 2013; Lei et al., 2019).

Hypertension

In spontaneously hypertensive rats, ischemic and pharmacological preconditioning confer benefits against ischemia-reperfusion injury (Boutros and Wang, 1995). Surprisingly, ischemic postconditioning failed to protect in spontaneously hypertensive rats, (Penna et al., 2010; Wagner et al., 2013), an effect that is attributed to the lack of phosphorylation of glycogen synthase kinase 3 (Gsk-3 β), as Gsk-3 β inhibitors restored the protection (Gonzalez Arbelaez et al., 2013). In contrast, postconditioning was successful in protecting rats with hypertensive dilated cardiomyopathy induced by administration of angiotensin II (Hernandez-Resendiz et al., 2013), an effect mediated by activation of Akt. Although remote ischemic and normobaric hypoxic conditioning can reduce blood pressure in hypertensive patients, there is no knowledge as to whether the SAFE pathway may be altered in the presence of hypertension (Lyamina et al., 2011; Madias, 2015).

In conclusion, the discovery of the SAFE path has brought a better understanding to the intrinsic prosurvival signaling cascade involved in conditioning-induced cardioprotection. Although the end targets of the SAFE pathway are still unknown, the discovery of this pathway has the potential to develop novel therapies/strategies to limit ischemia-reperfusion injuries by targeting some components of this pathway such as specific miRNAs, for example. It is important, however, to keep in mind that, if the activation of this pathway has been confirmed in small animals and pigs, its protective effect still needs to be confirmed in patients in which many co-morbidities and co-medications may affect its effectiveness.

Conflict of interest

None.

Acknowledgements

The research of SL is supported by the University of Cape Town and the National Research Foundation (111801), The Cancer Association of South Africa and Winetech. AI received a postdoctoral fellowship from the University of Cape Town and NH received a PhD fellowship from the Medical Research Council in South Africa.

References

- Adam T, Sharp S, Opie LH, Lecour S (2013) Loss of cardioprotection with ischemic preconditioning in aging hearts: role of sirtuin 1? J Cardiovasc Pharmacol Ther 18:46-53.
- Amantea D, Tassorelli C, Russo R, Petrelli F, Morrone LA, Bagetta G, Corasaniti MT (2011) Neuroprotection by leptin in a rat model of permanent cerebral ischemia: effects on STAT3 phosphorylation in discrete cells of the brain. Cell Death Dis 2:e238.
- Behmenburg F, Heinen A, Bruch LV, Hollmann MW, Huhn R (2017) Cardioprotection by Remote Ischemic Preconditioning is Blocked in the Aged Rat Heart in Vivo. J Cardiothorac Vasc Anesth 31:1223-1226.
- Boengler K, Hilfiker-Kleiner D, Heusch G, Schulz R (2010) Inhibition of permeability transition pore opening by mitochondrial STAT3 and its role in myocardial ischemia/ reperfusion. Basic Res Cardiol 105:771-785.
- Boengler K, Ungefug E, Heusch G, Schulz R (2013) The STAT3 inhibitor stattic impairs cardiomyocyte mitochondrial function through increased reactive oxygen species formation. Curr Pharm Des 19:6890-6895.
- Boengler K, Buechert A, Heinen Y, Roeskes C, Hilfiker-Kleiner D, Heusch G, Schulz R (2008) Cardioprotection by ischemic postconditioning is lost in aged and STAT3deficient mice. Circ Res 102:131-135.
- Boutros A, Wang J (1995) Ischemic preconditioning, adenosine and bethanechol protect spontaneously hypertensive isolated rat hearts. J Pharmacol Exp Ther 275:1148-1156.
- Brulhart-Meynet MC, Braunersreuther V, Brinck J, Montecucco F, Prost JC, Thomas A, Galan K, Pelli G, Pedretti S, Vuilleumier N, Mach F, Lecour S, James RW, Frias MA (2015) Improving reconstituted HDL composition for efficient post-ischemic reduction of ischemia reperfusion injury. PLoS One 10:e0119664.
- Cai D, Xaymardan M, Holm JM, Zheng J, Kizer JR, Edelberg JM (2003) Age-associated impairment in TNF-alpha

- Calvert JW, Jha S, Gundewar S, Elrod JW, Ramachandran A, Pattillo CB, Kevil CG, Lefer DJ (2009) Hydrogen sulfide mediates cardioprotection through Nrf2 signaling. Circ Res 105:365-374.
- Cohen MV, Baines CP, Downey JM (2000) Ischemic preconditioning: from adenosine receptor to KATP channel. Annu Rev Physiol 62:79-109.
- Cour M, Lecour S (2019) Remote ischaemic conditioning: in search of a suitable match. Nat Rev Cardiol 16:704-705.
- Deuchar GA, Opie LH, Lecour S (2007) TNFalpha is required to confer protection in an in vivo model of classical ischaemic preconditioning. Life Sci 80:1686-1691.
- Downey JM, Davis AM, Cohen MV (2007) Signaling pathways in ischemic preconditioning. Heart Fail Rev 12:181-188.
- Dube S, Matam T, Yen J, Mang HE, Dagher PC, Hato T, Sutton TA (2017) Endothelial STAT3 Modulates Protective Mechanisms in a Mouse Ischemia-Reperfusion Model of Acute Kidney Injury. J Immunol Res 2017:4609502.
- Ferdinandy P, Hausenloy DJ, Heusch G, Baxter GF, Schulz R (2014) Interaction of risk factors, comorbidities, and comedications with ischemia/reperfusion injury and cardioprotection by preconditioning, postconditioning, and remote conditioning. Pharmacol Rev 66:1142-1174.
- Frias MA, Lecour S, James RW, Pedretti S (2012) High density lipoprotein/sphingosine-1-phosphate-induced cardioprotection: Role of STAT3 as part of the SAFE pathway. JAKSTAT 1:92-100.
- Fuglesteg BN, Suleman N, Tiron C, Kanhema T, Lacerda L, Andreasen TV, Sack MN, Jonassen AK, Mjos OD, Opie LH, Lecour S (2008) Signal transducer and activator of transcription 3 is involved in the cardioprotective signalling pathway activated by insulin therapy at reperfusion. Basic Res Cardiol 103:444-453.
- Gonzalez Arbelaez LF, Perez Nunez IA, Mosca SM (2013) Gsk-3beta inhibitors mimic the cardioprotection mediated by ischemic pre- and postconditioning in hypertensive rats. Biomed Res Int 2013:317456.
- Goto M, Cohen MV, Downey JM (1996) The role of protein kinase C in ischemic preconditioning. Ann N Y Acad Sci 793:177-190.
- Han YS, Lee JH, Yoon YM, Yun CW, Noh H, Lee SH (2016) Hypoxia-induced expression of cellular prion protein improves the therapeutic potential of mesenchymal stem cells. Cell Death Dis 7:e2395.
- Hausenloy DJ et al. (2019) Effect of remote ischaemic conditioning on clinical outcomes in patients with acute myocardial infarction (CONDI-2/ERIC-PPCI): a single-blind randomised controlled trial. Lancet 394:1415-1424.
- Heinen A, Behmenburg F, Aytulun A, Dierkes M, Zerbin L, Kaisers W, Schaefer M, Meyer-Treschan T, Feit S, Bauer I, Hollmann MW, Huhn R (2018) The release of cardioprotective humoral factors after remote ischemic preconditioning in humans is age- and sex-dependent. J Transl Med 16:112.
- Hernandez-Resendiz S, Roldan FJ, Correa F, Martinez-Abundis E, Osorio-Valencia G, Ruiz-de-Jesus O, Alexanderson-Rosas E, Vigueras RM, Franco M, Zazueta C (2013) Postconditioning protects against reperfusion injury in hypertensive dilated cardiomyopathy by activating MEK/ ERK1/2 signaling. J Card Fail 19:135-146.
- Hu H, Nan J, Sun Y, Zhu D, Xiao C, Wang Y, Zhu L, Wu Y, Zhao J, Wu R, Chen J, Yu H, Hu X, Zhu W, Wang J (2017) Electron leak from NDUFA13 within mitochondrial complex I attenuates ischemia-reperfusion injury via dimerized STAT3. Proc Natl Acad Sci U S A 114:11908-11913.

- Kalakech H, Hibert P, Prunier-Mirebeau D, Tamareille S, Letournel F, Macchi L, Pinet F, Furber A, Prunier F (2014) RISK and SAFE signaling pathway involvement in apolipoprotein A-I-induced cardioprotection. PLoS One 9:e107950.
- Kurdi M, Booz GW (2007) Can the protective actions of JAK-STAT in the heart be exploited therapeutically? Parsing the regulation of interleukin-6-type cytokine signaling. J Cardiovasc Pharmacol 50:126-141.
- Kurrelmeyer KM, Michael LH, Baumgarten G, Taffet GE, Peschon JJ, Sivasubramanian N, Entman ML, Mann DL (2000) Endogenous tumor necrosis factor protects the adult cardiac myocyte against ischemic-induced apoptosis in a murine model of acute myocardial infarction. Proc Natl Acad Sci U S A 97:5456-5461.
- Kwan JC, Gao L, Macdonald PS, Hicks M (2015) Cardioprotective signalling by glyceryl trinitrate and cariporide in a model of donor heart preservation. Heart Lung Circ 24:306-318.
- Lacerda L, Somers S, Opie LH, Lecour S (2009) Ischaemic postconditioning protects against reperfusion injury via the SAFE pathway. Cardiovasc Res 84:201-208.
- Lamont KT, Somers S, Lacerda L, Opie LH, Lecour S (2011) Is red wine a SAFE sip away from cardioprotection? Mechanisms involved in resveratrol- and melatonininduced cardioprotection. J Pineal Res 50:374-380.
- Lecour S (2009a) Multiple protective pathways against reperfusion injury: a SAFE path without Aktion? J Mol Cell Cardiol 46:607-609.
- Lecour S (2009b) Activation of the protective Survivor Activating Factor Enhancement (SAFE) pathway against reperfusion injury: Does it go beyond the RISK pathway? J Mol Cell Cardiol 47:32-40.
- Lecour S, Rochette L, Opie L (2005a) Free radicals trigger TNF alpha-induced cardioprotection. Cardiovasc Res 65:239-243.
- Lecour S, Smith RM, Woodward B, Opie LH, Rochette L, Sack MN (2002) Identification of a novel role for sphingolipid signaling in TNF alpha and ischemic preconditioning mediated cardioprotection. J Mol Cell Cardiol 34:509-518.
- Lecour S, Suleman N, Deuchar GA, Somers S, Lacerda L, Huisamen B, Opie LH (2005b) Pharmacological preconditioning with tumor necrosis factor-alpha activates signal transducer and activator of transcription-3 at reperfusion without involving classic prosurvival kinases (Akt and extracellular signal-regulated kinase). Circulation 112:3911-3918.
- Lee SC, Jeong HJ, Lee SK, Kim SJ (2016) Hypoxic Conditioned Medium From Human Adipose-Derived Stem Cells Promotes Mouse Liver Regeneration Through JAK/STAT3 Signaling. Stem Cells Transl Med 5:816-825.
- Lei S, Su W, Xia ZY, Wang Y, Zhou L, Qiao S, Zhao B, Xia Z, Irwin MG (2019) Hyperglycemia-Induced Oxidative Stress Abrogates Remifentanil Preconditioning-Mediated Cardioprotection in Diabetic Rats by Impairing Caveolin-3-Modulated PI3K/Akt and JAK2/STAT3 Signaling. Oxid Med Cell Longev 2019:9836302.
- Lejay A, Fang F, John R, Van JA, Barr M, Thaveau F, Chakfe N, Geny B, Scholey JW (2016) Ischemia reperfusion injury, ischemic conditioning and diabetes mellitus. J Mol Cell Cardiol 91:11-22.
- Li H, Liu Z, Wang J, Wong GT, Cheung CW, Zhang L, Chen C, Xia Z, Irwin MG (2013) Susceptibility to myocardial ischemia reperfusion injury at early stage of type 1 diabetes in rats. Cardiovasc Diabetol 12:133.

Luo N, Liu J, Chen Y, Li H, Hu Z, Abbott GW (2018) Remote

ischemic preconditioning STAT3-dependently ameliorates pulmonary ischemia/reperfusion injury. PLoS One 13:e0196186.

- Lyamina NP, Lyamina SV, Senchiknin VN, Mallet RT, Downey HF, Manukhina EB (2011) Normobaric hypoxia conditioning reduces blood pressure and normalizes nitric oxide synthesis in patients with arterial hypertension. J Hypertens 29:2265-2272.
- Madias JE (2015) Sustained blood pressure lowering effect of twice daily remote ischemic conditioning sessions in a normotensive/prehypertensive subject. Int J Cardiol 182:392-394.
- Nan J, Hu H, Sun Y, Zhu L, Wang Y, Zhong Z, Zhao J, Zhang N, Wang Y, Wang Y, Ye J, Zhang L, Hu X, Zhu W, Wang J (2017) TNFR2 Stimulation Promotes Mitochondrial Fusion via Stat3- and NF-kB-Dependent Activation of OPA1 Expression. Circ Res 121:392-410.
- Niemi JP, DeFrancesco-Lisowitz A, Cregg JM, Howarth M, Zigmond RE (2016) Overexpression of the monocyte chemokine CCL2 in dorsal root ganglion neurons causes a conditioning-like increase in neurite outgrowth and does so via a STAT3 dependent mechanism. Exp Neurol 275 Pt 1:25-37.
- Ostadal B, Netuka I, Maly J, Besik J, Ostadalova I (2009) Gender differences in cardiac ischemic injury and protection--experimental aspects. Exp Biol Med (Maywood) 234:1011-1019.
- Paggio A, Checchetto V, Campo A, Menabo R, Di Marco G, Di Lisa F, Szabo I, Rizzuto R, De Stefani D (2019) Identification of an ATP-sensitive potassium channel in mitochondria. Nature 572:609-613.
- Park YH, Lee YM, Kim DS, Park J, Suk K, Kim JK, Han HS (2013) Hypothermia enhances induction of protective protein metallothionein under ischemia. J Neuroinflammation 10:21.
- Pedretti S, Brulhart-Meynet MC, Montecucco F, Lecour S, James RW, Frias MA (2019) HDL protects against myocardial ischemia reperfusion injury via miR-34b and miR-337 expression which requires STAT3. PLoS One 14:e0218432.
- Penna C, Tullio F, Moro F, Folino A, Merlino A, Pagliaro P (2010) Effects of a protocol of ischemic postconditioning and/or captopril in hearts of normotensive and hypertensive rats. Basic Res Cardiol 105:181-192.
- Penna C, Settanni F, Tullio F, Trovato L, Pagliaro P, Alloatti G, Ghigo E, Granata R (2013) GH-releasing hormone induces cardioprotection in isolated male rat heart via activation of RISK and SAFE pathways. Endocrinology 154:1624-1635.
- Penna C, Tullio F, Merlino A, Moro F, Raimondo S, Rastaldo R, Perrelli MG, Mancardi D, Pagliaro P (2009) Postconditioning cardioprotection against infarct size and post-ischemic systolic dysfunction is influenced by gender. Basic Res Cardiol 104:390-402.
- Przyklenk K, Maynard M, Greiner DL, Whittaker P (2011) Cardioprotection with postconditioning: loss of efficacy in murine models of type-2 and type-1 diabetes. Antioxid Redox Signal 14:781-790.
- Sakata H, Narasimhan P, Niizuma K, Maier CM, Wakai T, Chan PH (2012) Interleukin 6-preconditioned neural stem cells reduce ischaemic injury in stroke mice. Brain 135:3298-3310.
- Schulman D, Latchman DS, Yellon DM (2002) Urocortin protects the heart from reperfusion injury via upregulation of p42/p44 MAPK signaling pathway. Am J Physiol Heart Circ Physiol 283:H1481-1488.
- Smith RM, Suleman N, McCarthy J, Sack MN (2002) Classic

ischemic but not pharmacologic preconditioning is abrogated following genetic ablation of the TNFalpha gene. Cardiovasc Res 55:553-560.

- Somers SJ, Lacerda L, Opie L, Lecour S (2011) Age, genetic characteristics and number of cycles are critical factors to consider for successful protection of the murine heart with postconditioning. Physiol Res 60:971-974.
- Somers SJ, Frias M, Lacerda L, Opie LH, Lecour S (2012) Interplay between SAFE and RISK pathways in sphingosine-1-phosphate-induced cardioprotection. Cardiovasc Drugs Ther 26:227-237.
- Suleman N, Somers S, Smith R, Opie LH, Lecour SC (2008) Dual activation of STAT-3 and Akt is required during the trigger phase of ischaemic preconditioning. Cardiovasc Res 79:127-133.
- Sun XJ, Mao JR (2018) Role of Janus kinase 2/signal transducer and activator of transcription 3 signaling pathway in cardioprotection of exercise preconditioning. Eur Rev Med Pharmacol Sci 22:4975-4986.
- Tamareille S, Mateus V, Ghaboura N, Jeanneteau J, Croue A, Henrion D, Furber A, Prunier F (2011) RISK and SAFE signaling pathway interactions in remote limb ischemic perconditioning in combination with local ischemic postconditioning. Basic Res Cardiol 106:1329-1339.
- Tyagi S, Singh N, Virdi JK, Jaggi AS (2019) Diabetes abolish cardioprotective effects of remote ischemic conditioning: evidences and possible mechanisms. J Physiol Biochem 75:19-28.
- Wagner C, Ebner B, Tillack D, Strasser RH, Weinbrenner C (2013) Cardioprotection by ischemic postconditioning is abrogated in hypertrophied myocardium of spontaneously hypertensive rats. J Cardiovasc Pharmacol 61:35-41.

- Wang G, Zhou D, Wang C, Gao Y, Zhou Q, Qian G, DeCoster MA (2010) Hypoxic preconditioning suppresses group III secreted phospholipase A2-induced apoptosis via JAK2-STAT3 activation in cortical neurons. J Neurochem 114:1039-1048.
- Wang M, Tsai BM, Crisostomo PR, Meldrum DR (2006) Tumor necrosis factor receptor 1 signaling resistance in the female myocardium during ischemia. Circulation 114:I282-289.
- Wang M, Crisostomo PR, Markel TA, Wang Y, Meldrum DR (2008) Mechanisms of sex differences in TNFR2mediated cardioprotection. Circulation 118:S38-45.
- Wang M, Wang Y, Abarbanell A, Tan J, Weil B, Herrmann J, Meldrum DR (2009) Both endogenous and exogenous testosterone decrease myocardial STAT3 activation and SOCS3 expression after acute ischemia and reperfusion. Surgery 146:138-144.
- Watson AJ, Gao L, Sun L, Tsun J, Jabbour A, Ru Qiu M, Jansz PC, Hicks M, Macdonald PS (2013) Enhanced preservation of the rat heart after prolonged hypothermic ischemia with erythropoietin-supplemented Celsior solution. J Heart Lung Transplant 32:633-640.
- Yang G, Yang Y, Li Y, Hu Z (2020) Remote liver ischemic preconditioning protects rat brain against cerebral ischemia/reperfusion injury by activation AKT dependent pathway. Exp Physiol. 1-12.https://doi.org/10.1113/ EP088394